

**DETECTION OF *Burkholderia pseudomallei* USING POLYMERASE
CHAIN REACTION (PCR) IN SHEEP (*Ovis aries*)
IN NUEVA ECIJA**

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BIOGRAPHICAL SKETCH

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ABSTRACT

SAYCO, DENNIS, JR O., College of Veterinary Science and Medicine, Central Luzon State University, Science City of Muñoz, Nueva Ecija, Philippines, **June 2019**, **DETECTION OF *Burkholderia pseudomallei* USING POLYMERASE CHAIN REACTION (PCR) IN SHEEP (*Ovis aries*) IN NUEVA ECIJA**

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Melioidosis is a bacterial disease that affects humans and many species of animals including sheep. It is systemic disease that may affect almost any organ in an affected animal (Benoit et al., 2015) and a disease of public health importance in southeast Asia that is associated with high case-fatality rates in animals and humans (Cheng et al., 2005). The study was performed to detect *Burkholderia pseudomallei* from nasal swab of sheep around Nueva Ecija using Polymerase Chain Reaction/

A total of 40 sheep that are randomly picked at Nueva Ecija were collected with nasal swab samples. Extracted samples were subjected to bacterial culture and DNA extraction. Identification of *Burkholderia pseudomallei* was molecularly confirmed through Polymerase Chain Reaction with a primer of 16S rRNA gene and Tat domain gene primer and were subjected to gel analysis for identification.

Results showed that all 40 samples were negative from 16S rRNA gene and Tat domain gene primer that was used for the detection of *Burkholderia pseudomallei*.

Keywords: *Burkholderia pseudomallei*.; Melioidosis; Polymerase Chain Reaction; 16sr DNA gene; Tat domain gene primer

LITERATURE CITED

- Baker Al, Ezzahir J, Gardiner C, Shipton W, and Warner JM. (2015). Environmental attributes influencing the distribution of *Burkholderia pseudomallei* in Northern Australia. *PLoS One* 10(9):e0138953. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/26398904>
- Bauernfeind A, Roller C, Meyer D, Jungwirth R, and Schneider I. (1998). Molecular procedure for rapid detection of *Burkholderia mallei* and *Burkholderia pseudomallei*. *Journal of Clinical Microbiology* 36: 2737-41. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/9705426>
- Benoit TJ, Blaney DD, Doker TJ, Gee JE, Elrod MG, Rolim DB, Inglis TJ, Hoffmaster AR, Bower WA, and Walke HT. (2015). A review of melioidosis cases in the Americas. *American Journal of Tropical Medicine and Hygiene*. 93(6):1134-9. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4674224/>
- Bioterrorism Agents/Diseases. (2016). Retrieved from <https://emergency.cdc.gov/agent/agentlist-category.asp>
- Brett, P. J., Deshazer, D., and Woods, D. E. (1997). Characterization of *Burkholderia pseudomallei* and *Burkholderia pseudomallei*-like strains. *Epidemiol. Infect* 118: 137–148. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/9129590>
- Bureau of Animal Industry. (2018) Meat and Meat Products of 2017. Retrieved from <http://www.bai.da.gov.ph/index.php/stakeholders-corner/importation-data>
- Casas, N. (2018). Raising Sheep in the Philippines. 2017. Retrieved from <http://businessdiary.com.ph/430/raising-sheep-in-the-philippines/>
- Center for Disease Control and Prevention (CDC). (2006). Melioidosis. Retrieved from <http://www.cdc.gov.tw/uploads/files/03b6323f-a38c-4e43-9e12a7701e88db52.pdf>
- Chen, Y., Lin, H., Mu, J., Chiang, C., Chen, C., Buu, L., Lin, Y. E. and Chen, Y., (2010). Distribution of Melioidosis Cases and Viable *Burkholderia pseudomallei* in Soil: Evidence for Emerging Melioidosis in Taiwan. *Journal of Clinical Microbiology* 48 (4): 1432-1434. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2849618>
- Cheng, A. C. and Currie, B. J. (2005). Melioidosis: Epidemiology, Pathophysiology, and Management. *Clinical microbiology reviews* 18 (2): 383–416. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/15831829>

- Choy, J. L. (2017). Overview of Melioidosis. Retrieved from <https://www.msdsvetmanual.com/generalized-conditions/melioidosis/overview-of-melioidosis#v3273746>
- Dance, D. A. (2000). Ecology of *Burkholderia pseudomallei* and the interactions between environmental *Burkholderia* spp. and human-animal hosts. *Acta Trop* 74:159-168. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/10674645>
- Doker TJ, Quinn CL, Salehi ED, Sherwood JJ, Benoit TJ, Glass EM, GEE JE, Shadomy SV, Bower WA, Hoffmaster AR, Walke HT, Blaney DD, and Diorio MS. (2014). Melioidosis Investigation Team. Fatal *Burkholderia pseudomallei* infection initially reported as a *Bacillus* species. *American Journal of Tropical Medicine and Hygiene* 91(4):743-6. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/25092821>
- Egwuatu. TO., F. T. Ogunsola, I. M. Okodugha, B. Jide, D.G. Arewa, and O. A. Osinupebi. (2014). Effect of Blood Agar from Different Animal Blood on Growth Rates and Morphology of Common Pathogenic Bacteria. *Advances in Microbiology* 1238-1241. Retrieved from https://file.scirp.org/pdf/AiM_2014123014065959.pdf
- Engelthaler DM, Bowers J, Schupp JA, Pearson T, and Ginther J. (2011). Molecular investigations of a locally acquired case of melioidosis in southern AZ, USA. *PLoS Neglected Tropical Diseases* 5(10):e1347. Retrieved from <https://journals.plos.org/plosntds/article?id=10.1371/journal.pntd.0001347>
- Food and Agricultural Organization of United Nations (FAO). (2019). AGA NEWS: Economic analysis of animal diseases. Retrieved from http://www.fao.org/ag/againfo/home/en/news_archive/2016_Economic_analysis_animal_diseases.html
- Foong, Y. C., Tan, M., and Bradbury, R.S. (2014). Melioidosis: A Review. *Rural and Remote Health* 14: 2763. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/25359677>
- Gariyban, L. and Avashia, N. (2013). Research Techniques Made Simple: Polymerase Chain Reaction (PCR). *J Invest Dermatol.* 133(3). Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4102308/>
- Garner, G., Saville, P., and Fediavesky, A. (2003). Manual for the recognition of exotic diseases of livestock: A reference guide for animal health staff. Food and Agriculture Organization of the United Nation (FAO). Retrieved from <http://www.spc.int/rahs>

- Gee JE, Sacchi CT, Glass MB, De BK, Weyant RS, Levett PN, Whitney AM, Hoffmaster AR, and Popovic T. (2003) Use of 16S rRNA gene sequencing for rapid identification and differentiation of *Burkholderia pseudomallei* and *B. mallei*. *J Clin Microbiol.* 2003 Oct;41(10):4647-54. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/14532197>
- Haase, A. Brennan, M., Barrett, S., Wood, Y., Huffam, S. O'brien, D., and Currie, B. (1998). Evaluation of PCR for Diagnosis of Melioidosis. *Journal of clinical microbiology* 36 (4): 1039–1041. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC104685/>
- Hagen, R. M., Gauthier, Y. P., Sprague, L. D., Vidal, D. R., Zysk, G., Finke, E-J., and Neubauer, H. (2002). Technical Report: Strategies for PCR based detection of *Burkholderia pseudomallei* DNA in paraffin wax embedded tissues. *Journal of Clinical Pathology: Molecular Pathology* 55:398–400. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1187279/>
- Hall CM, Busch JD, Shippy K, Allender CJ, Kaestli M, Mayo M, Sahl JW, Schupp JM, Colman RE, Keim P, Currie BJ, and Wagner DM. (2015). Diverse *Burkholderia* species isolated from soils in the southern United States with no evidence of *B. pseudomallei*. *PLoS One* 10(11):e0143254. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/26600238>
- Hampton V, Kaestli M, Mayo M, Choy JL, Harrington G, Richardson L, Benedict S, Noske R, Garnett ST, Godoy D, Spratt BG, and Currie BJ. (2011). Melioidosis in birds and *Burkholderia pseudomallei* dispersal, Australia. *Emerging Infectious Diseases* 17(7):1310-2. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3381411/>
- Haque, Q. M. (2010). Evaluation of Available Diagnostic Tests for Melioidosis. *Journal of Taibah University Medical Sciences* 5(2): 89-97. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/29720430>
- Heymann DL., (2006). Melioidosis. *Control of Communicable Diseases Manual Edition.* 18: 386-8 Retrieved from <https://www.navybmr.com/study%20material/CCDM.pdf>
- Hillsborough Community College (2018). BLOOD AGAR NOTES. Retrieved from <https://www.hccfl.edu/media/580776/blood%20agar%20notes.pdf>
- Ho C., Lau C., Martelli P., Chan S., Tse C., Wu A., Yuen K., Lau SK., and Woo P. (2011). Novel Pan-Genomic Analysis in Target Selection for Multiplex PCR Identification and Detection of *Burkholderia pseudomallei*, *Burkholderia thailandensis*, and

Burkholderia cepacia Complex Species: A Proof of Concept Study. *Journal of Clinical Microbiology*. 49(3):814-821. Doi: 10.1128/JCM.01702-10. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3067743/>

Howard, K. and Inglis, T. J. J. (2003). Novel Selective Medium for Isolation of *Burkholderia pseudomallei*. *Journal of Clinical Microbiology* 41 (7): 3312–3316. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC165331>

Inglis, T. J. J. (2010). The Treatment of Melioidosis. *Pharmaceuticals* 3: 1296-1303. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4033981>

James GL, Delaney B, Ward L, Freeman K, Mayo M, and Currie BJ. (2013). Surprisingly low seroprevalence of *Burkholderia pseudomallei* in exposed healthy adults in the Darwin region of tropical Australia where melioidosis is highly endemic. *Clinical Vaccine Immunology* 20(5):759-60. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3647750>

Kaestli M, Harrington G, Mayo M, Chatfield MD, Harrington I, Hill A, Munksgaard N, Gibb K, Currie BJ. (2015). What drives the occurrence of the melioidosis bacterium *Burkholderia pseudomallei* in domestic gardens? *PLoS Neglected Tropical Diseases* 9(3):e0003635. Retrieved from <https://journals.plos.org/plosntds/article?id=10.1371/journal.pntd.0003635>

Lau SK, Sridhar S, Ho CC, Chow WN, Lee KC, Lam CW, Yuen KY and Woo PC. (2015). Laboratory diagnosis of melioidosis: past, present and future. *Experimental Biology and Med (Maywood)*. 2015;240(6):742-51. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/25908634>

Latoza, R. (2015). WV records highest goat, sheep production in PH. Retrieved: <http://www.iloilometropolitantimes.com/wv-records-highest-goat-sheep-production-in-ph/>

Lazar A, N. R., Govan, B., Cullinane, M., Harper, M., Adler, B., and Boyce, J. D. (2009). The molecular and cellular basis of pathogenesis in melioidosis : how does *Burkholderia pseudomallei* cause disease?. *FEMS Microbiology Review* 33:1079–1099. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/19732156>

Lee, N., J. L. Wu, C. H. Lee, and W. C. Tsai. (1985). *Pseudomonas pseudomallei* infection from drowning: the first reported case in Taiwan. *Journal of Clinical Microbiology* 22:352-354. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC268408/>

- Limmathurotsakul, D., and Peacock, S. J. (2011). Melioidosis: A clinical Overview. *British Medical Bulletin* 99: 125–139. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/21558159>
- Millan JM, Mayo M, Gal D, Janmaat A, and Currie BJ. (2007) Clinical variation in melioidosis in pigs with clonal infection following possible environmental contamination from bore water. *Veterinary Journal* 174:200-2. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/16807011>
- Mohanty, S., Pradhan, G., Panigrahi, M. K., Mohapatra, P. R., and Mishra, B. (2016). A case of systemic melioidosis: unravelling the etiology of chronic unexplained fever with multiple presentations. *Pneumonol Alergol Pol* 84: 121–125. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/27238172>
- Montúfar Fe, Ochoa Je, Ortega H, Franco L, Montúfar Mc, Monsalve A, Jaramillo C, and Zapata M. (2015). Melioidosis in Antioquia, Colombia: an emerging or endemic disease? A case series. *International Journal of Infectious Diseases* 37:50-7. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/26051974>
- Peacock, S. J., Chieng, G., Cheng, A. C., Dance, D. A. B., Amornchai, P., Wongsuvan, G., Teerawattanasook, N., Chierkul, W., Day, N. P. J. Raja NS, Ahmed MZ, and Singh NN. (2005). Melioidosis: an emerging infectious disease. *Journal of Postgraduate Medicine* 51:140-5. Retrieved from <http://www.jpgmonline.com/article.asp?issn=0022-3859;year=2005;volume=51;issue=2;spage=140;epage=145;aulast=Raja>
- Philippine Council for Agriculture, Aquatic, and Natural Resources Research and Development (PCAARRD) (2011). Goat/Sheep. Retrieved from http://www.pcaarrd.dost.gov.ph/home/momentum/ruminants/index.php?option=com_content&task=view&id=131&Itemid=175
- Philippines Statistics Authority (PSA). (2018). Goat Situation Report, January - June 2018. Retrieved from <https://psa.gov.ph/livestock-poultry-ipsr/goat/inventory>
- Rega, P. P. (2015). CBRNE – Glanders and Melioidosis. Retrieved from <https://emedicine.medscape.com/article/830235-overview>
- Samy, R. P., Stiles, B. G., Sethi, G., and Lim, L. H. K. (2017). Melioidosis: Clinical impact and public health threat in the tropics. *Plos: neglected tropical diseases*. 1-28. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/28493905>
- Solanki, G. (2012). Polymerase Chain Reaction. *International Journal of Pharmacologica Research* 2(3): 98 – 101. Retrieved from

https://www.researchgate.net/publication/271729269_Polymerase_Chain_Reaction

acio, H. D. (2017). Raising Small Ruminants for food and profit. Retrieved from <http://edgedavao.net/agri-trends/2017/01/18/raising-small-ruminants-food-profit/>

ne Swedish University of Agricultural Sciences (SLU). (2016). *Burkholderia pseudomallei*. Retrieved from <http://www.vetbact.org/?artid=49>

rakulsomboon, S., V. Vuddhakul, P. Tharavichitkul, N. Na-gnam, Y. Suputtamongkol and V. Thamlikitkul. (1999). Epidemiology of arabinose assimilation in *Burkholderia pseudomallei* isolated from patients and soil in Thailand. *Southeast Asian Journal of Tropical Medical Public Health* 30:756-759. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/10928371>

Vuthiekanun, V. (2005). Comparison of Ashdown's Medium, *Burkholderia cepacia* Medium, and *Burkholderia pseudomallei* Selective Agar for Clinical Isolation of *Burkholderia pseudomallei*. *Journal of Clinical Microbiology* 43 (10): 5359–5361. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC1248505/>

Vuthiekanun, V. and Dance, D. (2012). Standard Operating Procedure (SOP): Simplified Method for the Isolation of *Burkholderia pseudomallei* from soil 3-4. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC3264119/>

Tabuchi E., Kosako, Y., Oyaizu H., Oyaizu, H., Yano, I., Hotta, H., Hashimoto, Y., Ezaki T., and Arakawa, M. (1992). Proposal of *Burkholderia* gen. nov. and transfer of seven species of the genus *Pseudomonas* homology group II to the new genus, with the group species *Burkholderia cepacia* (Palleroni and Holmes 1981) comb. nov. *Journal of Clinical Microbiology*, 36(12), 1251-1275. Retrieved from <https://www.ncbi.nlm.nih.gov/pubmed/1283774>