

**GROWTH AND SURVIVAL OF SNAKEHEAD (*Channa* sp.) LARVAE FED
WITH BRINE SHRIMP (*Artemia* sp.) NAUPLII AND *Moina* sp.**

by

JEROME JACOB QUIDASOL

An Undergraduate Thesis presented to the faculty of the College of Fisheries In partial fulfillment of the requirements for the degree of

BACHELOR OF SCIENCE IN FISHERIES

**Department of Aquatic Post-Harvest
COLLEGE OF FISHERIES
CENTRAL LUZON STATE UNIVERSITY
Science City of Munoz, Nueva Ecija**

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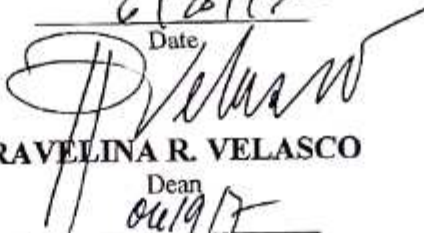
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GROWTH AND SURVIVAL OF SNAKEHEAD (*Channa* sp.) LARVAE FED WITH BRINE SHRIMP (*Artemia* sp.) NAUPLII AND *Moina* sp.^{1/}

ABSTRACT

Two experiments were conducted with the objective of assessing the effect of two selected diets on the growth performance, survival rate and food preference of snakehead fish larvae (*Channa* sp.). The experiments were carried out using 60-liters capacity glass aquaria with 20 liters of water volume. Live food such as *Artemia* sp. nauplii and *Moina* sp. were given to larvae three times daily from 1-21 days after hatching when the yolk sac was totally absorbed by the larvae. The first experiment was conducted to evaluate the growth performance and survival rate of snakehead larvae fed with *Artemia* sp. nauplii and *Moina* sp. Results showed higher length gain, weight gain, and SGR in larvae fed with *Artemia* sp. nauplii compared to larvae fed with *Moina* sp. with values 23.3187 mm, 118.35 mg, 11.4994 % and 21.7714 mm, 84.4267 mg, 10.0124 %, respectively. In the present study, higher mortality due to cannibalism was also observed in larvae fed with *Artemia* sp. nauplii (19.6667 ± 6.50) compared to *Moina* sp. (14.3333 ± 10.04). Meanwhile, survival rate was observed to be higher in larvae fed with *Moina* sp. compared to larvae fed with *Artemia* sp. nauplii with values, 56.3333 ± 10.59 and 45 ± 15.04 , respectively. Statistically, no significant difference was observed in both treatments in terms of length gain, weight gain, SGR, cannibalism and survival rate. The second experiment aimed to determine the food preference of snakehead fish (*Channa* sp.) larvae. It was found out that the snakehead fish age 13, 20 and 27 days old consumed both *Artemia* sp. nauplii and *Moina* sp. However, statistical analysis showed significantly higher relative percentage occurrence of *Moina* sp. compared to *Artemia* sp. nauplii in the digestive tract of snakehead fish of age 13, 20 and 27 days old with values 65.6218, 67.0354, 54.6904 % and 34.3782, 32.9646 and 45.3096 %, respectively. The study suggested that *Moina* sp. is a suitable live food and a potential substitute for *Artemia* sp. nauplii in the larval rearing of snakehead fish (*Channa* sp.).

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- Boonyaratpalin, M., E.W. McCoy and T. Chittapalpong. 1985. Snakehead culture and its socio-economics in Thailand. Annual Report. Fisheries and Aquaculture Department, Thailand. Retrieved on October 11, 2016 from <http://www.fao.org>.
- Boyd, C.E. 1982. Water quality management of pond fish culture. Elsevier Sci. Publ. Co. New York, USA. 318 p.
- Buskey, E.J. 2005. Behavioral characteristics of copepods that affect their suitability as food for larval fishes. p. 91-105. In: Lee, C.S., P.J. O'Bryen and N.H. Marcus (eds.). Copepods in Aquaculture. Blackwell Publishing. Iowa, USA. 269 p.
- Dave, G. 1989. Experiences with waste water cultured *Daphnia* in the start feeding of rainbow trout (*Salmo gairdneri*). Aquaculture, 79:337-343.
- Dayal, R., P.P. Srivastava, A. Bhatnagar, S. Chowdhary, A.K. Yadav and W.S. Lakra. 2012. Comparative utilization impact of various dietary lipids on growth indices in striped mullet, *Channa striatus* (Bloch) fingerlings. Journal of Animal and Feed Research, 1: 64-69.
- Diana, J.S., W.Y.B. Chang, D.R. Ottey and W. Chuapohuk. 1985. Production systems for commonly cultured freshwater fishes of Southeast Asia. International Program Report. No. 7. Great Lake of Marine Water Center, University of Michigan, Ann Arbor, MI, pp. 75-79.
- Evangelista, A.D., N.R. Fortes and C.B. Santiago. 2005. Comparison of some live organisms and artificial diet as feed for Asian catfish *Clarias macrocephalus* (Gunther) larvae. J. Appl. Ichthyol., 21: 437-443.
- Faruque, M.M., M.D. Kawser ahmed and M.M.A. Quddus. 2010. Use of live food and artificial diet supply for the growth and survival of African catfish (*Clarias gariepinus*). World J. Zoo., 5: 82-89.
- Fermin, A.C. and M.E.C. Bolivar. 1991. Larval rearing of the Philippine freshwater catfish, *Clarias macrocephalus* (Gunther) fed live zooplankton and artificial diet: a preliminary study. Bamigdgeh, 43: 87-94.
- Fermin, A.C. and R.D. Recometa. 1988. Larval rearing of bighead carp, *Aristichthys nobilis* Richardson, using different types of feed and their combinations. Aquaculture and Fisheries Manage., 19: 283-290.
- Govoni, J.J., Boehlert, G.W. and Y. Watanabe. 1986. The physiology of digestion of fish larvae. Environ. Biolo. Fishes, 16:59-77.
- Hetch, T. and S. Appelbaum. 1987. Note on the growth of Israeli sharptooth catfish (*Clarias gariepinus*) during the primary nursing phase. Aquaculture 63: 195-204.

- Higgs, D.A., J.R. Market, M.D. Plonikoff, J. R McBride and M. Dosanjh. 1985. Development of nutritional and environmental strategies for maximizing the growth and survival of juvenile pink salmon (*Oncorhynchus gribuscha*). *Aquaculture*, 47: 113-130.
- Hoff, F.H. and T.W. Snell. 1989. Plankton culture manual. Florida Aquafarms Inc: Pisces Publ. Group Deven.CT. Florida, 125 pp.
- Hogendoorn, H. 1980. Controlled propagation of the African catfish, *Clarias lazera* (C. & V.) III. Feeding and growth of fry. *Aquaculture*, 21: 233-241.
- Jais, A.M.M., R. McCulloch and K. Croft. 1994. Fatty acid and amino acid composition in haruan as a potential role in wound healing. *Gen. Pharmacol.*, 25: 947-950.
- James, R., J. Muthukrishnan, K. Sampath. 1993. Effects of food quality on temporal and energetics cost of feeding in *Cyprinus carpio* (Cyprinidae). *J. Aquac. Trop.* 8:47-53.
- Kahan, D. 1984. Overview: food for larval stages in aquaculture live or artificial food? *Israel J. Zool.*, 3: 152-153.
- Kanazawa, A., S. Teshima and K. Ono. 1979. Relationship between essential fatty acid requirements of aquatic animals and the capacity for bioconversion of linoleic acid to highly unsaturated fatty acids. *Comp. Biochem. Physiol.*, 63: 295-298.
- Knud-Hansen, C.F., T.R. Batterson, C.D. McNabb, Y. Hadiroseyani, D. Dana and H. Muhammed Eidman. 1990. Hatchery techniques for egg and fry production of *Clarias batrachus* (Linnaeus). *Aquaculture*, 89: 9-19.
- Kumar, D., K. Marimuthu, M.A. Haniffa and T.A. Sethuramalingam. 2008. Effect of different live feed on growth and survival of stripped murrel *Channa striatus* larvae. *Fish. Aquatic Sci.*, 25: 105-110.
- Lauff, M. and R. Hofer. 1984. Proteolytic enzymes in fish developed and the importance of dietary enzymes. *Aquaculture*, 37: 335-346.
- Lavens, P. and P. Sorgeloos. 1996. Manual on the production and use of live food for Aquaculture. FAO, Fisheries Technical Paper no. 361. Romjpe. FAO, 295: 283-288.
- Léger, P., D.A. Bengtson, K.L. Simpson and P. Sorgeloos. 1986. The use and nutritional value of *Artemia* as a food source. *Oceanogr. Mar.Biol. Annu., Rev.* 24: 521-623.

- Ling, S.W. 1977. Snakehead, *Ophiocephalus* spp. culture. p. 60-66. In: Ling S.W. (ed.). Aquaculture in Southeast Asia: A historical overview. University of Washington Press, Seattle. 108 p.
- Marimuthu, K. and M.A. Haniffa. 2006. Larval rearing of the spotted snakehead *Channa punctatus* fed with different feeding regimes. J. Fish. Aquatic. Sci., 1(3): 297-300.
- Marimuthu, K. and M.A. Haniffa. 2007. Embryonic and larval development of the striped snakehead *Channa striatus*. Taiwania, 52: 84-92.
- Marimuthu, K., M.A. Haniffa, M. Muruganandam and A.J. Arockia Raj. 2001a. Low cost murrel seed production technique for fish farmer. Naga. 24 (1&2): 21-22.
- _____. 2001b. Spawning and parental behavior in the induced bred murrels. Indian Journal of Fisheries, 48(4): 409-411.
- Marioghae, I.E. 1982. Notes on the biology and distribution of *M. vollenhovenii*, *M. macrobrachion* in the Lagos lagoon. Rev. Zool. Afr., 96(3): 493-508.
- Mat Jais, A.M., R. McCulloch and K. Croft. 1994. Fatty acid and amino acid composition in Haruan as a potential role in wound healing. Gen. Pharmacol., 25:947-950.
- Mehraj Ud Din, W., K. Altaff and M. A. Haniffa. 2009. Study on the growth and survival of *Channa striatus* (Bloch) post-larvae using live feed. Bangladesh J. Fish. Res., 13: 131-135.
- Mgaya, Y.D. and J.P. Mercer. 1995. The effect of size grading and stocking density on the growth performance of juvenile abalone, *Haliotis tuberculata* Linn. Aquaculture. 136:297-312.
- Mollah, M.F.A., M.S.A. Mamum, M.N. Sarowar and A. Roy. 2009. Effect of stocking density on the growth and breeding performance of brood fish and larval growth and survival of shol, *Channa striatus* (Bloch). Journal of Bangladesh Agriculture University, 7(2): 427-432.
- Muntaziana, M.P.A., S.M.N. Amin, M. Aminur Rahman, A.A. Rahim and K. Marimuthu. 2013. Present culture status of the endangered snakehead, *Channa striatus* (Bloch, 1793). Asian J. Ani. Vet. Adv., 8: 369-375.
- Ng, P.K.L. and K.K.P. Lim. 1990. Snakeheads (Pisces: *Channidae*): natural history biology and economics importance. p. 127-152. In: Chou, L.M. and K.L.P. Ng (eds.). Essays in Zoology. Papers Commemorating the 40th Anniversary of the Department of Zoology. National University of Singapore, Singapore. Retrieved on October 11, 2016 from <http://www.snakeheads.org>.

- Parameswaran, S. and V.K. Murugesan. 1976. Observation on the hypophysation of murrels (*Ophiocephalidae*). *Hydrobiologia*, 50 (1): 81-87.
- Punia, P. 1988. Culture of *Moina micrura* on various organic waste products. *Journal of the Indian Fisheries Association*, 18: 129-134.
- Qin, J. and A.W. Fast. 1997. Food selection and growth of young snakehead *Channa striatus*. *J. Appl. Ichthyol.*, 13: 12-25.
- Qin, J., Fast, A.W., D. DeAnda and R.P. Weidenbach. 1997. Growth and survival of larval snakehead fish (*Channa striatus*) fed different diets. *Aquaculture*. 148: 105-113.
- Qin, J.G. and A.W. Fast. 1998. Effect of temperature, size and density on culture performance of snakehead *Channa striatus* (Bloch), fed formulated feed. *Aquacult. Res.*, 29: 299-303.
- Rahim, M.H.A., A. Rozila and A.M.M. Jais. 2009. The physical-chemical and morphological study of huruan (*Channa striatus*) in peninsular Malaysia. *Res. Biol. Sci.*, 4: 994-1009.
- Rottmann, R.W., Graves, S.J., Watson, C. and Roy, P. E. Yanong. 2003. Culture techniques of *Moina*: The ideal Daphnia for feeding freshwater fish fry. CIR 1054, FAO, Rome: 2-9.
- Sampath, K. 1985. Food intake, conversion and surfacing activity as a function of density and water change frequency in an air-breathing fish, *Channa striatus*. *Aquaculture*, 46: 201-213.
- Sarowar, M.N., M.Z.H. Jewel, M.A. Sayeed and M.F.A. Mollah. 2010. Impact of different diets on growth and survival of *Channa striatus* fry. *Int. J. Biol. Res.*, 1: 8-12.
- Sorgeloos, P. 1980. The use of the brine shrimp *Artemia* in Aquaculture. p. 25-46. *In*: Persoone, G., P. Sorgeloos, O. Roels and E. Jaspers (Editors). *The Brine Shrimp Artemia: Ecology, Culturing, and Use in Aquaculture*. Vol. 3. Universa Press. Belgium. 456 p.
- Tawaratmanikul, P., T. Viputanimat, A. Mewan and K. Pokasap. 1988. Study on the suitable *Moina* density nursing the giant catfish, *Pangasionodon gigas*. Technical paper no. 6/1988, Thailand: Pathumthani freshwater fisheries station, Inland fisheries division, Department of Fisheries, Ministry of Agriculture and Cooperatives, 6 p. (in Thai with English abstract).

- Villegas, C.T. 1990. The effect on growth and survival of feeding water flees (*Moina macrocopa*) and rotifer (*Branchionus plicatilis*) to milkfish (*Chanos chanos*) fry. *Bamidgeh*, 42: 10-17.
- War, M., K. Altaff and M.A. Haniffa. 2011. Growth and survival of larval snakehead *Channa stratus* (Bloch, 1793) fed different live feed organisms. *Turkish Journal of Fisheries and Aquatic Science*, 11: 523-528.
- Watanabe, T., C. Kitajima and S. Fujita. 1983. Nutritional values of live organisms used in Japan for mass propagation of fish: a review. *Aquaculture*, 34: 115-143.
- Wee, K.L. 1982. Snakeheads –their biology and culture. p. 181-213. *In*: Muir, R. (ed.). *Recent Advances in Aquaculture*. Westview Press. Boulder, Co. 453 p.

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