

ENRICHMENT OF PHENOLIC ANTIOXIDANT OF SABA BANANA (*Musa acuminata x balbisiana*) PEEL BY SOLID-STATE BIOCONVERSION USING *Rhizopus oryzae*

EULYNNE CHIOCO EUGENIO



An Undergraduate Thesis Submitted to the Faculty of the Department of Chemistry,
College of Arts and Sciences, Central Luzon State University,
Science City of Muñoz, Nueva Ecija, Philippines
in partial fulfillment of the requirements
for the degree of

BACHELOR OF SCIENCE IN CHEMISTRY

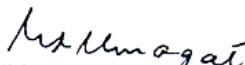
August 2017

CHE-03-17-001


APPROVAL AND ACCEPTANCE

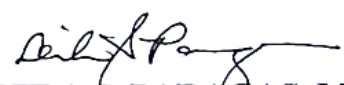
The undergraduate thesis entitled **ENRICHMENT OF PHENOLIC ANTIOXIDANT OF SABA BANANA** (*Musa acuminata x balbisiana*) **BY SOLID-STATE BIOCONVERSION USING** *Rhizopus oryzae* prepared and submitted by **EULYNNE C. EUGENIO** in partial fulfillment of the requirements for the degree of **BACHELOR OF SCIENCE IN CHEMISTRY** is hereby

APPROVED:



MYRNA R. UMAGAT, Ph.D.
Adviser
09-08-17
Date

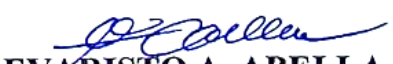

ROSALIE R. RAFAEL, Ph.D.
Critic
09-08-17
Date


REDEL L. GUTIERREZ, Ph.D.
Department Research Coordinator
09-08-17
Date


DANILO S. PARAGAS, M.S.
Department Chair
09-08-17
Date

AND ACCEPTED:


RICH MILTON R. DULAY, M.S.
College Research Coordinator
09-11-17
Date


EVARISTO A. ABELLA, Ph.D.
Acting Dean
09-11-17
Date

BIOGRAPHICAL SKETCH

PERSONAL INFORMATION

Name : **EULYNNE CHIOCO EUGENIO**
Date of Birth : October 8, 1996
Place of Birth : Quezon City

EDUCATIONAL BACKGROUND

Elementary : **Bettbien Montessori**
Malasin, San Jose City, Nueva Ecija
March 2008
Secondary : **Central Luzon State University Science High School**
Science City of Muñoz, Nueva Ecija
April 2012
Tertiary : **Central Luzon State University**
Science City of Muñoz, Nueva Ecija
Bachelor of Science in Chemistry
August 2017

SCHOLARSHIP AND AWARDS

Dean's List : 1st sem and 2nd sem 2012-2013; 1st sem 2013-2014
College Scholar : 1st sem 2016-2017

SEMINARS ATTENDED

Participant : 2013 PhilRice Nutrition Month Seminar
Central Experiment Station, PhilRice Institute
Science City of Muñoz, Nueva Ecija
July 30, 2013
: Shimadzu Technology for Analytical Instrumentation
CAS Little Theater, CLSU
October 08, 2015

: National Chemistry Student's Congress 2017
NISMED Auditorium, University of the
Philippines, Diliman, Quezon City
March 04, 2017

ACKNOWLEDGMENT

I am so grateful to God for always giving me wisdom, strength and hope in times of doubt. I failed more than once doing this study, but God gave me more than enough to finish what I have started. I cannot finish this study without His works in my life and His words in my heart.

I am also thankful to all the people who are the sources of all to finish this study:

To my family, especially my parents, Mrs. Jonelyn C. Eugenio and Ladislao E. Eugenio Jr., for their faith in me. They provided all of my needs, and their sacrifices are the source of my strength.

To my adviser, Dr. Myrna R. Umagat, for her time, patience, kindness, advises and knowledge that she had imparted to improve the study. I really appreciate her in giving time even on her busy days doing her other responsibilities in the university.

To my critic, Dr. Rosalie R. Rafael, for allowing me to conduct my thesis in the Biochemical Organic and Natural Products (BONP), her shared knowledge about natural products, and her suggestions to improve my paper.

To Prof. Danila S. Paragas, Chemistry Department Chair, for the guidance during the conduct of research especially during overnights.

To Dr. Redel L. Gutierrez, Department Research Coordinator, for his patience and concern during the study.

To Diana C. Castillo for sharing her knowledge on microorganisms, and letting me use their laboratory. She had helped me to find the right method on the inoculation.

To Sarah Joy Antonio, Regine Cate Saul, and Eddyson Rafael for their assistance during the conduct of my thesis at BONP. Ate Sarah guided me on DPPH radical

scavenging assay, Ate Cate on media preparation, and Kuya Eddyson on the use of autoclave.

To my classmates, Lei Ann, Jenina, Mary Ann, Alison, Diolina, Mikka, Leendel, Erahem, and Jay-Lord, for their motivations, support and happiness they had shared during our laboratory works.

To Keen Reyes, for his assistance during the research, his patience, kindness and understanding, and his words of encouragement. He stayed and lifted me up during the darkest days of my life.

Lastly, I am also to all the people who contributed to the success of this research. We did it! To God be all the glory!

EULYNNE C. EUGENIO

TABLE OF CONTENTS

	Page
TITLE PAGE	i
APPROVAL AND ACCEPTANCE	ii
BIOGRAPHICAL SKETCH	iii
ACKNOWLEDGEMENT	v
LIST OF TABLES	ix
LIST OF FIGURES	x
LIST OF APPENDIX TABLES	xi
LIST OF APPENDIX FIGURES	xii
ABSTRACT	1
1 INTRODUCTION	
1.1 Background of the Study	2
1.2 Significance of the Study	3
1.3 Objectives of the Study	4
1.4 Scope and Limitations of the Study	5
1.5 Time and Place of the Study	5
2 REVIEW OF RELATED LITERATURE	
2.1 Description	6
2.2 Musaceae Family	7
2.3 Saba Banana	7
2.4 Antioxidant Activity	9
2.5 Phenolic Compounds in Banana Peels	9
2.6 Solid-State Bioconversion	11
2.7 Microorganism	13
2.8 Related Studies	14
3 METHODOLOGY	
3.1 Chemical and Reagents	17

3.2	Microorganism	17
3.3	Substrate	18
3.4	Media and Cultivation	18
3.5	Crude Enzyme Extraction	28
3.6	Determination of Total Phenolic Content	19
3.7	DPPH Radical Scavenging Activity	19
3.8	Hydrogen Peroxide Scavenging Assay	20
3.9	Statistical Analyses	
4	RESULTS AND DISCUSSION	
4.1	Total Phenolic Content of Saba banana peel with and without <i>Rhizopus oryzae</i>	21
4.2	Antioxidant Activity of Saba banana peel with and without <i>Rhizopus oryzae</i>	27
4.2.1	DPPH Radical Scavenging Activity	27
4.2.2	Hydrogen Peroxide Radical Scavenging Activity	30
5	SUMMARY, CONCLUSIONS, AND RECOMMENDATION	
5.1	Summary	32
5.2	Conclusions	33
5.3	Recommendations	34
	LITERATURE CITED	35
	APPENDICES	41

LIST OF TABLES

Table No.	Title	Page
4.1	Comparison of Total Phenolic Content (mg/GAE g) of Saba banana peel with and without <i>Rhizopus oryzae</i> between the treatments	21
4.2	Comparison of Total Phenolic Content (mg/GAE g) of Saba banana peel with and without <i>Rhizopus oryzae</i> within the days of fermentation	22
4.3	Comparison of Total Phenolic Content (mg/GAE g) of Saba banana peel with and without <i>Rhizopus oryzae</i>	26
4.4	Comparison of DPPH radical scavenging activity (%) of Saba banana peel with and without <i>Rhizopus oryzae</i> between the treatments	27
4.5	Comparison of DPPH radical scavenging activity (%) of Saba banana peel with and without <i>Rhizopus oryzae</i> within the days of fermentation	28
4.6	Comparison of DPPH radical scavenging activity (%) of Saba banana peel with and without <i>Rhizopus oryzae</i>	30

LIST OF FIGURES

Figure No.	Title	Page
2.1	Saba banana (<i>Musa acuminata x balbisiana</i>)	8
4.1	Total Phenolic Content (mg/ GAE g) of Saba banana peel with and without <i>Rhizopus oryzae</i> plotted against the days of fermentation	24
4.2	DPPH radical scavenging (%) of Saba banana peel with and without <i>Rhizopus oryzae</i> plotted against the days of fermentation	29

LIST OF APPENDIX TABLE

Appendix Table No.	Title	Page
4.1	Comparison of H ₂ O ₂ radical scavenging activity (%) of Saba banana peel with and without <i>Rhizopus oryzae</i> between the treatment	45
4.2	Comparison of H ₂ O ₂ radical scavenging (%) of Saba banana peel with and without <i>Rhizopus oryzae</i>	45
5.1	Statistical Tests of Total Phenolic content per days of fermentation	47
5.2	Statistical Tests of Total Phenolic content between treatments	48
5.3	Statistical Test of antioxidant activity per days of fermentation	50
5.4	Statistical Test of antioxidant activity between treatments	51

LIST OF APPENDIX FIGURES

Appendix Figure No.	Title	Page
1.1	Schematic Diagram	42
2.1	Calibration curve of gallic acid in the determination of total phenolic content (TPC) using Folin-Ciocalteu method.	43
3.1	Residue of Saba banana peel and Saba banana peel with <i>Rhizopus oryzae</i>	44
4.1	% H ₂ O ₂ radical scavenging of Saba banana peel with and without <i>Rhizopus oryzae</i> plotted against the days of fermentation	46
5.1	Statistical test plot of DPPH radical scavenging assay	49
5.2	Statistical test plot of H ₂ O ₂ radical scavenging assay	50

ENRICHMENT OF PHENOLIC ANTIOXIDANT OF SABA BANANA (*Musa acuminata x balbisiana*) BY SOLID-STATE BIOCONVERSION USING *Rhizopus oryzae*¹

EULYNNE C. EUGENIO

ABSTRACT

Background: Saba (*Musa acuminata x balbisiana*) banana is one of the common varieties of banana in the Philippines. Banana peels are wastes that contribute pollution to the environment, but also a good source of antioxidants. Solid-state bioconversion (SSB) was used to enrich the phenolic capacity of fruit wastes. *Rhizopus oryzae*, the fungi responsible for fermentation during SSB, can produce β -glucosidase enzyme that can free the bound phenolics during enzyme hydrolysis, and improve antioxidant activity. **Methods:** Two treatments were prepared: one (T1) containing Saba banana peel only, and the other (T2) containing Saba banana peel with 8mm *Rhizopus oryzae*. Treatments were fermented for 16 days. Both treatments were analyzed every after two days. The treatments were extracted after 48 hours of soaking using 80% ethanol. The filtrates were measured spectrophotometrically. The total phenolic content was measured using Folin-Ciocalteu method, and the antioxidant activity using DPPH and H₂O₂ radical scavenging activities. **Results:** The total phenolic content of Saba banana peel without *Rhizopus oryzae* is 0.45 mg/GAE g higher than Saba banana peel with *Rhizopus oryzae*. While in its antioxidant activity, Saba banana peel with *Rhizopus oryzae* is 1.61% higher in inhibiting DPPH radicals compared to Saba banana peel without *Rhizopus oryzae*. In the inhibition of H₂O₂ radicals, the obtained values were negative due to the extraction solvent used, ethanol. The radicals oxidized the ethanol, instead of being scavenged by the Saba banana peel extracts. **Conclusions:** SSB using *Rhizopus oryzae* cannot enrich the total phenolic content of Saba banana peel, and its antioxidant activity.

Keywords: solid-state bioconversion (SSB), *Rhizopus oryzae*, Saba banana (*Musa acuminata x balbisiana*), β -glucosidase

¹ An undergraduate thesis presented to the Faculty of the Department of Chemistry, College of Arts and Sciences, Central Luzon State University, Science City of Muñoz, Nueva Ecija in partial fulfillment of the requirements for the degree of Bachelor of Science in Chemistry. Prepared under the supervision of Professor Dr. Myrna R. Umagat and Professor Dr. Rosalie R. Rafael. August 2017. **CHE-03-17-001**

LITERATURE CITED

- Abdel-Hameed, E.-S. S. (2009). Total phenolic contents and free radical scavenging activity of certain Egyptian Ficus species leaf samples. *Food chemistry*, 114(4), 1271-1277.
- Abedinifar, S., Karimi, K., Khanahmadi, M., & Taherzadeh, M. J. (2009). Ethanol production by *Mucor indicus* and *Rhizopus oryzae* from rice straw by separate hydrolysis and fermentation. *Biomass and bioenergy*, 33(5), 828-833.
- Abrashid, S., Ibrahim, D., & Omar, I. C. (2011). Utilization of palm kernel cake for the production of mannanase by an indigenous filamentous fungus, *Aspergillus niger* USM F4 under solid substrate fermentation. *The Internet Journal of Microbiology*, 9(1), 1-19.
- Alam, M. Z., Elgharbawy, A. A., & Hamzah, M. (2013). *Optimization and characterization of Candida cylindracea lipase produced from palm kernel cake by solid-state bioconversion*. Paper presented at the AIChE Annual Meeting. San Francisco, USA.
- Azmi, N. A. B. (2010). *Extraction of antioxidant activity, phenolic content and minerals in banana peel*. Universiti Malaysia Pahang.
- Battaglia, E., Benoit, I., van den Brink, J., Wiebenga, A., Coutinho, P. M., Henrissat, B., & de Vries, R. P. (2011). Carbohydrate-active enzymes from the zygomycete fungus *Rhizopus oryzae*: a highly specialized approach to carbohydrate degradation depicted at genome level. *Bmc Genomics*, 12(1), 38.
- Beckman, C. H. (2000). Phenolic-storing cells: keys to programmed cell death and periderm formation in wilt disease resistance and in general defence responses in plants? *Physiological and Molecular Plant Pathology*, 57(3), 101-110.
- Bhanja, T., Kumari, A., & Banerjee, R. (2009). Enrichment of phenolics and free radical scavenging property of wheat koji prepared with two filamentous fungi. *Bioresource technology*, 100(11), 2861-2866.
- Bond, O. (2015). What Are the Benefits of Eating Banana Peels?
- Bravo, L. (1998). Polyphenols: chemistry, dietary sources, metabolism, and nutritional significance. *Nutrition reviews*, 56(11), 317-333.
- Buckley, P. M., Sommer, N., & Matsumoto, T. (1968). Ultrastructural details in germinating sporangiospores of *Rhizopus stolonifer* and *Rhizopus arrhizus*. *Journal of bacteriology*, 95(6), 2365-2373.
- Cantabrana, I., Perise, R., & Hernández, I. (2015). Uses of *Rhizopus oryzae* in the kitchen. *International Journal of Gastronomy and Food Science*, 2(2), 103-111. doi:<https://doi.org/10.1016/j.ijgfs.2015.01.001>

- Chang, Y., & Hudson, H. (1967). The fungi of wheat straw compost: I. Ecological studies. *Transactions of the British Mycological Society*, 50(4), 649-666.
- Correia, R., McCue, P., Magalhaes, M., Macedo, G., & Shetty, K. (2004). Phenolic Antioxidant Enrichment of Soy Flour-Supplemented Guava Waste by *Rhizopus oligosporus*-mediated Solid-State Bioprocessing. *Process Biochemistry*, 39, 2167-2172.
- Dela Cruz, F., Gueco, L., Damasco, O., Huelgas, V., Dela Cueva, F., Dizon, T., . . . Molina, A. (2008). Farmers' Handbook on Introduced and Local Banana Cultivars in the Philippines. *Bioversity International*, 78.
- Du, G., Li, M., Ma, F., & Liang, D. (2009). Antioxidant capacity and the relationship with polyphenol and vitamin C in Actinidia fruits. *Food chemistry*, 113(2), 557-562.
- Fatemeh, S., Saifullah, R., Abbas, F., & Azhar, M. (2012). Total phenolics, flavonoids and antioxidant activity of banana pulp and peel flours: influence of variety and stage of ripeness. *International Food Research Journal*, 19(3), 1041-1046.
- Ghribi, D., Abdelkefi-Mesrati, L., Mnif, I., Kammoun, R., Ayadi, I., Saadaoui, I., . . . Chaabouni-Ellouze, S. (2011). Investigation of Antimicrobial Activity and Statistical Optimization of *Bacillus subtilis* SPB1 Biosurfactant Production in Solid-State Fermentation. *Journal of Biomedicine and Biotechnology*. doi:10.1155/2012/373682
- Giasson, B. I., Ischiropoulos, H., Lee, V. M.-Y., & Trojanowski, J. Q. (2002). The relationship between oxidative/nitrative stress and pathological inclusions in Alzheimer's and Parkinson's diseases 1, 2. *Free Radical Biology and Medicine*, 32(12), 1264-1275.
- Goldstein, J. L., & Swain, T. (1963). Changes in tannins in ripening fruits. *Phytochemistry*, 2(4), 371-383.
- González-Montelongo, R., Lobo, G., & González, M. (2010). Antioxidant activity in banana peel extracts: testing extraction conditions and related bioactive compounds. *Food chemistry*, 119(3), 1030-1039.
- Gryganskyi, A. P., Lee, S. C., Litvintseva, A. P., Smith, M. E., Bonito, G., Porter, T. M., . . . Vilgalys, R. (2010). Structure, function, and phylogeny of the mating locus in the *Rhizopus oryzae* complex. *PLoS One*, 5(12), e15273.
- Guia, J., (2015). Common Tropical Fruit in the Philippines. Retrieved from <https://www.vigattintourism.com/tourism/articles/Common-Tropical-Fruits-in-the-Philippines>.
- Hardisson, A., Rubio, C., Baez, A., Martin, M., Alvarez, R., & Diaz, E. (2001). Mineral composition of the banana (*Musa acuminata*) from the island of Tenerife. *Food chemistry*, 73(2), 153-161.
- He, Y., Ma, X., & Lu, M. (2012). Oxidation of alcohols with hydrogen peroxide in the presence of a new triple-site phosphotungstate. 187-197.

- Hölker, U., & Lenz, J. (2005). Solid-state fermentation—are there any biotechnological advantages? *Current Opinion in Microbiology*, 8(3), 301-306.
- Reece J.B., Lisa A. Urry, M. C., Cain, M. L., Wasserman, S. A., Minorsky, P. V., & Jackson, R. B. (2014). *Campbell Biology* (10th ed.): Pearson.
- Jeong, S. H., Jung, J. Y., Lee, S. H., Jin, H. M., & Jeon, C. O. (2013). Microbial succession and metabolite changes during fermentation of dongchimi, traditional Korean watery kimchi. *International journal of food microbiology*, 164(1), 46-53.
- Kanazawa, K., & Sakakibara, H. (2000). High content of dopamine, a strong antioxidant, in cavendish banana. *Journal of Agricultural and Food Chemistry*, 48(3), 844-848.
- Kar, B., Banerjee, R., & Bhattacharyya, B. (1999). Microbial production of gallic acid by modified solid state fermentation. *Journal of industrial microbiology & biotechnology*, 23(3), 173-177.
- Karmakar, M., & Ray, R. (2011). A Statistical Approach for Optimization of Simultaneous Production of β -Glucosidase and Endoglucanase by *Rhizopus oryzae* from Solid-State Fermentation of Water Hyacinth Using Central Composite Design. *Biotechnology Research International*, 2011.
- Kawasaki, B. T., Hurt, E. M., Mistree, T., & Farrar, W. L. (2008). Targeting cancer stem cells with phytochemicals. *Molecular interventions*, 8(4), 174.
- Kinsey, B. (2017). Musaceae – Banana Family. Retrieved from <http://wildlifeofhawaii.com/flowers/category/plant-family/musaceae-banana-family/>
- Knapp, F., & Nicholas, H. (1969). The sterols and triterpenes of banana peel. *Phytochemistry*, 8(1), 207-214.
- Krisch, J., Takó, M., Papp, T., & Vágvölgyi, C. (2010). Characteristics and potential use of β -glucosidases from Zygomycetes. *Current research, technology and education topics in applied microbiology and microbial biotechnology*, 2, 891-896.
- Leterme, P., Buldgen, A., Estrada, F., & Londoño, A. M. (2006). Mineral content of tropical fruits and unconventional foods of the Andes and the rain forest of Colombia. *Food chemistry*, 95(4), 644-652.
- Lim, Y., Lim, T., & Tee, J. (2007). Antioxidant properties of several tropical fruits: A comparative study. *Food chemistry*, 103(3), 1003-1008.
- Mccue, P., Horii, A., & Shetty, K. (2003). Solid-State Bioconversion of Phenolic Antioxidants from defatted Soybean Powders by *Rhizopus oligosporus*: Role of Carbohydrate-Cleaving Enzymes *Journal of food biochemistry*, 27(6), 501-514.
- Mokbel, M. S., & Hashinaga, F. (2005). Antibacterial and antioxidant activities of banana (Musa, AAA cv. Cavendish) fruits peel. *American Journal of Biochemistry and Biotechnology*, 1(3), 125-131.

- Morton, J. (1987). Banana. *Fruits of warm climate*, 29-46.
- Mukherjee, G., & Banerjee, R. (2004). Biosynthesis of tannase and gallic acid from tannin rich substrates by *Rhizopus oryzae* and *Aspergillus foetidus*. *Journal of basic microbiology*, 44(1), 42-48.
- Mupure, C., & Chitindingu, K. (2006). Antioxidant potentials and degrees of polymerization of six wild fruits. *Scientific Research and Essays*, 1(3), 087-092.
- Musa, K. H., Abdullah, A., Jusoh, K., & Subramaniam, V. (2011). Antioxidant activity of pink-flesh guava (*Psidium guajava* L.): effect of extraction techniques and solvents. *Food Analytical Methods*, 4(1), 100-107.
- Navabi, S. M., Ebrahimzadeh, M. A., Navabi, A. F., Bahramian, F., & Bekhradnia, A. R. (2010). Antioxidant and Free Radical Scavenging Activity of *H. officinalis* L. var. *angustifolius*, *V. odorata*, *B. hyrcana* and *C. speciosum*. *Pakistan Journal of Pharmaceutical Sciences*, 23(1), 29-34.
- Nguyen, T. B. T., Ketsa, S., & van Doorn, W. G. (2003). Relationship between browning and the activities of polyphenoloxidase and phenylalanine ammonia lyase in banana peel during low temperature storage. *Postharvest Biology and Technology*, 30(2), 187-193.
- Okole, B., & Schulz, F. (1996). Micro-cross sections of banana and plantains (*Musa* spp.): morphogenesis and regeneration of callus and shoot buds. *Plant Science*, 116(2), 185-195.
- Palsdottir, H. (2016). Bananas: Good or Bad?. Retrieved from <http://www.healthline.com/nutrition/bananas-good-or-bad>
- Pantig, J. (2014). The Effects of Food Waste on the Environment. Retrieved from <https://donatedontdump.net/2014/07/07/the-effects-of-food-waste-on-the-environment-by-junemy-pantig/>
- Petruzzello, M. (2014). *Rhizopus*. *Encyclopedia Britannica*. Retrieved from <https://www.britannica.com/science/Rhizopus>
- Prakash, A., Rigelhof, F., & Miller, E. (2016). Antioxidant Activity. *Medallion Laboratories*.
- Rebello, L., Ramos, A., Petruzatti, P., Barcia, M., Munoz N., & Gutierrez, I (2013). Flour of banana (*Musa AAA*) peel as a source of antioxidant phenolic compounds. *Food Research International*, 55, 397-403.
- Robinson, J.C. 1996. Bananas and plantains. Wallingford, UK:CABI Publishing, CAB International.
- Rudrappa, U. (2016). Banana fruit nutrition facts.
- Sandhu, D., & Sidhu, M. (1980). The fungal succession on decomposing sugar cane bagasse. *Transactions of the British Mycological Society*, 75(2), 281-286.

- Santos, M. d., Pereira, E., Badiale, E., & Souza, L. d. (2012). Phenolic compounds and antioxidant activity in fermented rice (*Oryza sativa*) bran. 32(3).
- Sathya, M. (2014). Assaying the Antioxidant Activity of Banana Peel. *American Journal of Biochemistry and Molecular Biology*, 4: 122, 129.
- Seymour, G., Taylor, J., & Gregory, A. (1993). *Biochemistry of Fruit Ripening*.
- Shahidi, F., & Yeo, J. (2016). Insoluble-Bound Phenolics in Food. *Molecules*, 21.
- Singhania, R. R., Patel, A. K., Sukumaran, R. K., Larroche, C., & Pandey, A. (2013). Role and significance of beta-glucosidases in the hydrolysis of cellulose for bioethanol production. *Bioresource technology*, 127, 500-507.
- Someya, S., Yoshiki, Y., & Okubo, K. (2002). Antioxidant compounds from bananas (*Musa Cavendish*). *Food chemistry*, 79(3), 351-354.
- Stanton, R. (2016). Antioxidants: their role in health.
- Stover T.H., Simmonds N.W. 1987. Classification of banana cultivars. In: Stover RH, Simmonds NW, editors. Bananas. New York: Wiley. 97-103.
- Subagio, A., Morita, N., & Sawada, S. (1996). Carotenoids and their fatty-acid esters in banana peel. *Journal of nutritional science and vitaminology*, 42(6), 553-566.
- Sugiharto, S., Yudiarti, T., & Isroli, I. (2016). Assay of antioxidant potential of two filamentous fungi isolated from the Indonesian fermented dried cassava. *Antioxidants*, 5(1), 6.
- Suja, K., Jayalekshmy, A., & Arumughan, C. (2005). Antioxidant activity of sesame cake extract. *Food chemistry*, 91(2), 213-219.
- Sulaiman, S. F., Yusoff, N. A. M., Eldeen, I. M., Seow, E. M., Sajak, A. A. B., Supriatno, & Ooi, K. L. (2011). Correlation between total phenolic and mineral contents with antioxidant activity of eight Malaysian bananas (*Musa sp.*). *Journal of Food Composition and Analysis*, 24(1), 1-10. doi:<https://doi.org/10.1016/j.jfca.2010.04.005>
- Takii, Y., Ikeda, K., Sato, C., Yano, M., Sato, T., & Konno, H. (2005). Production and characterization of β -glucosidase from *Rhizopus oryzae* MIBA348. *J. Biol. Macromol*, 5(1), 11-16.
- Tang, Y., Zhang, B., Li, X., Chen, P. X., Zhang, H., Liu, R., & Tsao, R. (2016). Bound Phenolics of Quinoa Seeds Released by Acid, Alkaline, and Enzymatic Treatments and Their Antioxidant and α -Glucosidase and Pancreatic Lipase Inhibitory Effects. *Journal of Agricultural and Food Chemistry*, 64(8), 1712-1719. doi:10.1021/acs.jafc.5b05761
- Temanel, B. (2007). Techno-Guide for Saba Banana Production in Cagayan Valley. Retrieved from <http://ww1.openacademy.ph/>
- Thompson, J. (2014). Banana in the Philippines : Saba Banana, Known Also As Cardaba.

- Vattem, D. A., & Shetty, K. (2002). Solid-state production of phenolic antioxidants from cranberry pomace by *Rhizopus oligosporus*. *Food Biotechnology*, 16(3), 189-210.
- Wall, M. M. (2006). Ascorbic acid, vitamin A, and mineral composition of banana (*Musa* sp.) and papaya (*Carica papaya*) cultivars grown in Hawaii. *Journal of Food Composition and Analysis*, 19(5), 434-445.
- Ware, M. (2017). Bananas: Health benefits, facts, research.
- Xiao, Z., Zhang, X., Gregg, D. J., & Saddler, J. N. (2004). Effects of sugar inhibition on cellulases and β -glucosidase during enzymatic hydrolysis of softwood substrates. *Applied Biochemistry and Biotechnology*, 115(1), 1115-1126. doi:10.1385/abab:115:1-3:1115