



**CENTRAL LUZON STATE UNIVERSITY**



**VIABILITY EVALUATION OF EXTENDED POST-MORTEM  
GOAT EPIDIDYMAL SPERMATOZOA STORED AT  
REFRIGERATED CONDITION**

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An Undergraduate Thesis Submitted to the Faculty of the Department of  
Biological Sciences, College of Arts and Sciences, Central Luzon  
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**BACHELOR OF SCIENCE IN BIOLOGY**

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*-Philippians 4:13*

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ABSTRACT

**TOMAS, JULIUS V.** Bachelor of Science in Biology, Department of Biological Sciences, College of Arts and Sciences, Central Luzon State University, Science City of Munoz, Nueva Ecija, Philippines, June 2017. **VIABILITY EVALUATION OF EXTENDED POST-MORTEM GOAT EPIDIDYMAL SPERM STORED AT REFRIGERATED CONDITION.**

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**Co-Adviser: Lerma C. Ocampo, DVM, PhD**

The study was conducted to assess the viability of postmortem goat epididymal spermatozoa (EPS) recovered immediately in Tris-citric acid-lactose-raffinose (TCLR) buffer solution and transported at ambient temperature in the laboratory. Specific sperm quality parameters such as initial motility, sperm concentration, morphology (live/dead and normal/abnormal), pellet volume and pH of the buffer solution were determined. The pelletized epididymal sperm were diluted in TCLR with 20% egg yolk (v/v) and 7% glycerol (v/v) and stored in a refrigerator. The processed sperm were observed every 12 hours for sperm motility assessed by conventional method and computer assisted sperm analyzer (CASA) equipment. Representative samples were smear-stained to determine sperm livability and sperm morphology.

The recovered epididymal spermatozoa obtained a mean sperm concentration value of  $339 \pm 83.5$  and volume of  $(0.71 \pm 0.08)$ . There was a significant decline



( $p < 0.05$ ) in sperm motility from the time it was immediately recovered ( $50.6 \pm 5.14$ ), after it was processed 12 hpm ( $18.1 \pm 5.66$ ) and 24 hpm ( $5.94 \pm 1.19$ ) of storage in the refrigerator.. Similar observations of the total motility (CASA-MOT) and progressive motility (CASA-PMOT) values by CASA evaluation declined ( $p < 0.05$ ) considerably after 12 hpm (CASA-MOT- $12.05 \pm 6.20$ , CASA-PMOT- $5.64 \pm 3.50$ ) and 24 hpm (CASA-MOT- $7.02 \pm 7.57$ , CASA-PMOT- $2.43 \pm 2.69$ ) as compared with the freshly recovered sperm at 0 hpm (CASA-MOT- $45.98 \pm 8.34$ , CASA-PMOT- $20.84 \pm 4.99$ ).

Microscopic observations of live and dead epididymal spermatozoa registered significantly different mean values ( $p < 0.05$ ) that were inversely proportionally with each other at 0hpm (live- $74.10 \pm 5.39$ , dead- $25.51 \pm 5.42$ ), 12hpm (live- $45.09 \pm 9.69$ , dead- $54.11 \pm 10.04$ ) and 24hpm (live- $9.45 \pm 8.37$ , dead- $90.56 \pm 8.37$ ). The proportion of normal spermatozoa significantly ( $p < 0.05$ ) declined over time at 0 hpm ( $67.15 \pm 4.14$ ), 12 hpm ( $48.58 \pm 11.37$ ) and 24 hpm ( $9.04 \pm 8.08$ ). The incidence of sperm abnormalities significantly increased ( $p < 0.05$ ) in number as the storage time at  $0-5^{\circ}\text{C}$  was prolonged (0 hpm:  $32.79 \pm 4.15$ ), (12 hpm:  $51.69 \pm 10.99$ ) and (24 hpm:  $90.96 \pm 8.08$ ). Similar observation for the CASA abnormal epididymal spermatozoa considerably increased ( $p < 0.05$ ) upon reaching 12 hpm ( $96.33 \pm 2.03$ ) and 24 hpm ( $97.46 \pm 2.32$ ) of refrigerated storage condition. Some sperm abnormalities noted include cytoplasmic droplets (distal and proximal), bent tails, bent midpiece, coiled tail, large head, micro head, dag-like defect, distal reflex and detached head. Cytoplasmic droplets ranked the highest in both CASA and conventional methods of evaluation for sperm abnormality.



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