

**THE EFFECT OF LIGHT AND DARK CONFINEMENT ON THE GLUCOSE  
CONTENT, SKIN PIGMENTATION AND SURVIVAL RATE  
OF RED TILAPIA (*Oreochromis* spp.)**

**By**

**ABIGAIL CONRADO CAVITA**

**An Undergraduate Thesis presented to faculty of the College of Fisheries in partial  
fulfilment of the requirements for the degree of**

**BACHELOR OF SCIENCE IN FISHERIES**

**COLLEGE OF FISHERIES  
CENTRAL LUZON STATE UNIVERSITY  
SCIENCE CITY OF MUÑOZ, NUEVA ECIJA  
PHILIPPINES**

**2017**



**COLLEGE OF FISHERIES  
CENTRAL LUZON STATE UNIVERSITY  
Science City of Muñoz, Nueva Ecija**

**THE EFFECT OF LIGHT AND DARK CONFINEMENT IN THE GLUCOSE  
CONTENT, SKIN PIGMENTATION AND SURVIVAL RATE  
OF RED TILAPIA (*Oreochromis* spp.)**

By

**ABIGAIL C. CAVITA**

Undergraduate thesis presented to the Faculty of the  
College of Fisheries in partial fulfilment of the  
Requirements for the degree of

**BACHELOR OF SCIENCE IN FISHERIES**

**APPROVED:**

**RAVELINA R. VELASCO**

Adviser

\_\_\_\_\_ Date

  

**KARL MARX A. QUIAZON**

Critic

\_\_\_\_\_ Date

**NOTED:**

  

**REMEDIOS B. BOLIVAR**

Department Chair

\_\_\_\_\_ Date

**RECORDED:**

  

**REMEDIOS B. BOLIVAR**  
College Research Coordinator

\_\_\_\_\_ Date

**ACCEPTED:**

  

**RAVELINA R. VELASCO**

Acting Dean

\_\_\_\_\_ Date

## BIOGRAPHICAL DATA



### PERSONAL DATA

Name	Abigail C. Cavita
Birthday	January 22, 1997
Birthplace	Loob, Gainza, Camarines Sur
Address	111 Purok Kayumanggi Loob, Gainza, Camarines Sur
Parents	Mr. Jaime S.B. Cavita Mrs. Agripina C. Cavita

### EDUCATIONAL ATTAINMENT

Elementary	Loob Elementary School Loob, Gainza, Camarines Sur
Secondary	Gainza National High School San Juan, Gainza, Camarines Sur
Tertiary	Central Luzon State University Science City of Muñoz, Nueva Ecija

## ACKNOWLEDGEMENTS

The author would like to take this opportunity to give gratitude to those people who gave their help, knowledge, support, encouragement, guidance and valuable time for accomplishment of this paper.

First of all, to ALMIGHTY GOD for the wisdom, blessings and for the love He gave. For providing things the author needed to accomplish this paper.

To the author's parents, Mrs. Agripina Cavita and Mr. Jaime Cavita for the love and for supporting every decision the author made. Thank you for all the things you've done, for making the author of what she is today. To her siblings: Jinna, Victor, Christian and Jason for the support.

To Dr. Ravelina R. Velasco, the author's adviser, for the constructive criticism, sharing knowledge, guidance and encouragement for the accomplishment of this paper.

To Dr. Karl Marx A. Quiazon, the author's critic, for sharing his knowledge, time and for motivating the author to finish this paper.

To Dr. Remedios B. Bolivar, the College Research Coordinator, for the patience in giving comments and criticism to improve this paper.

To all the faculty and staff of College of Fisheries for the lessons learned that is essential to make this study possible.

To Engr. Zaldy Bartolome for lending the author the fish needed to conduct her study; to Ma'am Juliet Holasca, Kuya Ruel, Ate Ave joy and Ma'am Grace for their help and assistance during the conduct of this study.

To Bureau of Fisheries and Aquatic Resources for the scholarship grant and for helping the author financially.

To her friends: Jan, Rein, Jeph, Josh, Joni, Aphrodite, Carlo, Geneva, Xyrra, Daniel, April, Cellyne, Kae and Monique for the help, assistance, encouragement and for the friendship that the author will treasure the most.

To her classmates for the shared moments and making memorable the author stays in CLSU.

To Thames Jeuss Alexis Reñosa for patience, love, encouragement and helping the author during the conduct of this study.

**ABIGAIL CONRADO CAVITA**

## TABLE OF CONTENTS

	<b>Page</b>
<b>LIST OF TABLES</b>	vii
<b>LIST OF FIGURES</b>	viii
<b>LIST OF APPENDIX TABLE</b>	ix
<b>LIST OF APPENDIX FIGURE</b>	xii
<b>ABSTRACT</b>	xii
<b>INTRODUCTION</b>	
Background of the Study	1
Statement of the Problem	3
Significance of the Study	3
Objective of the Study	4
Scope and Limitation	4
Time and Place of the Study	4
<b>REVIEW OF RELATED LITTERATURE</b>	
Red tilapia	5
Light and dark confinement	5
Glucose content	7
Skin pigment	8
<b>MATERIALS AND METHODS</b>	
Experimental fish	9
Experimental design	9
Experimental treatments	10
Experimental layout	10
Data Gathered	11
Statistical analysis	13

## **RESULTS AND DISCUSSION**

Glucose content in the blood of Red tilapia	14
Skin coloration	15
Growth performance	18
Water Quality parameters	20

<b>SUMMARY, RECOMMENDATION AND CONCLUSION</b>	26
---	----

<b>LITERATURE CITED</b>	28
-------------------------	----

<b>APPENDICES</b>	34
-------------------	----

## LIST OF TABLES

<b>Table No.</b>	<b>Title</b>	<b>Page</b>
1	Treatments and description of the variables evaluated	10
2	Mean blood glucose values ( $\pm$ SD) obtained from different background color	14
3	Mean skin color values obtained from light and dark confinement	17
4	Mean growth performance of Red tilapia throughout the duration of the study	18

## LIST OF FIGURES

<b>Figure No.</b>	<b>Title</b>	<b>Page</b>
1	Experimental layout	10
2	Glucometer used in measuring blood glucose	11
3	Skin color chart for Red tilapia	12
4a	Mean dissolved oxygen measured in light and dark confinement	21
4b	Mean temperature measured in light and dark confinement	21
4c	Mean pH measured in light and dark confinement	22

## LIST OF APPENDIX TABLES

<b>Appendix Table No.</b>	<b>Title</b>	<b>Page</b>
1	Initial and final bulk weight of fish prior to the experiment	35
2	Mortality of fish recorded prior to the experiment	35
3	Blood glucose level measured during experiment	36
4	Skin color examined during experiment	37
5	Mean dissolved oxygen monitored during experiment	38
6	Mean temperature monitored during experiment	39
7	Mean pH monitored during experiment	40
8	Analysis of variance of dissolved oxygen in the morning during the experiment	41
9	Comparison of mean dissolved oxygen in the morning during the experiment	41
10	Analysis of variance of dissolved oxygen in the afternoon during the experiment	41
11	Comparison of mean dissolved oxygen in the afternoon during the experiment	42
12	Analysis of variance of temperature in the morning during the experiment	42
13	Comparison of mean temperature in the morning during the experiment	42
14	Analysis of variance of temperature in the afternoon during the experiment	43
15	Comparison of mean temperature in the morning during the experiment	43
16	Analysis of variance of pH in the morning during the experiment	43
17	Comparison of mean pH in the morning during the experiment	44

18	Analysis of variance of pH in the afternoon during the experiment	44
19	Comparison of mean pH in the afternoon during the experiment	44
20	Analysis of variance of the survival during the experiment	45
21	Comparison of mean of the survival during the experiment	45
22	Analysis of variance of the initial color in dorsal portion during the experiment	45
23	Comparison of mean of the initial color in dorsal portion during the experiment	46
24	Analysis of variance of the final color in dorsal portion during the experiment	46
25	Comparison of mean of the final color in dorsal portion during the experiment	46
26	Analysis of variance of the initial color in caudal portion during the experiment	47
27	Comparison of mean of the initial color in caudal portion during the experiment	47
28	Analysis of variance of the final color in caudal portion during the experiment	47
29	Comparison of mean of the final color in caudal portion during the experiment	48
30	Analysis of variance of the initial color in belly portion during the experiment	48
31	Comparison of mean of the initial color in belly portion during the experiment	48
32	Analysis of variance of the final color in belly portion during the experiment	49
33	Comparison of mean of the final color in belly portion during the experiment	49

34	Analysis of variance of the initial glucose content during the experiment	49
35	Comparison of mean of the initial glucose content during the experiment	50
36	Analysis of variance of the final glucose content during the experiment	50
37	Comparison of mean of the final glucose content during the experiment	50
38	Analysis of variance of the initial weight during the experiment	51
39	Comparison of mean of the initial weight during the experiment	51
40	Analysis of variance of the final weight during the experiment	51
41	Comparison of mean of the final weight during the experiment	52
42	Analysis of variance of SGR during the experiment	52
43	Comparison of mean of SGR during the experiment	52
44	Analysis of variance of BGW during the experiment	53
45	Comparison of mean of BGW during the experiment	53

## LIST OF APPENDIX FIGURES

<b>Table No.</b>	<b>Title</b>	<b>Page</b>
1	Experimental set- up	54
2	Monitoring of water temperature, dissolved oxygen and pH	54
3	Fish cultured in no black cover aquaria	55
4	Fish cultured in black covered aquaria but no top cover	55
5	Fish cultured in black covered aquaria with top cover	56
6	Collecting blood sample from a fish	56
7	Reading of blood glucose	57

**THE EFFECT OF LIGHT AND DARK CONFINEMENT ON THE GLUCOSE  
CONTENT, SKIN PIGMENTATION AND SURVIVAL RATE  
OF RED TILAPIA (*Oreochromis spp.*)<sup>1/</sup>**

**ABSTRACT**

This study aimed to evaluate the effect of light and dark confinement in the glucose content, skin pigmentation and survival rate of Red tilapia (*Oreochromis spp.*). Three treatments were used in this study namely the control (no black plastic cover), T<sub>2</sub> (covered with black plastic without top cover and plastic) and T<sub>3</sub> (covered with black plastic with top cover. In all the blood measurements, the blood glucose ranged from low to medium (34.11±2.51 to 74.53±16.46). No significant differences were found in glucose content. For skin coloration, the dorsal, caudal and belly portions were considered for comparison. Using the improvised color chart with intensity of 1 (light pink) to 13 (orange), T<sub>2</sub> had the highest color intensity. In terms of growth, the SGR was highest in T<sub>2</sub>. For weight gain, T<sub>1</sub> obtained the highest. The mean survival rate was highest in T<sub>1</sub> and lowest in T<sub>3</sub>. The low survival that was determined may be due to handling, fish injection and absence of light. In conclusion, this study revealed that light and dark confinement significantly affects the color of red tilapia. T<sub>2</sub> showed the best color of red tilapia as exhibited by its orange color. The blood glucose content was not affected by light and dark confinement.

---

<sup>1/</sup> Undergraduate thesis presented to the faculty of College of Fisheries, Central Luzon State University as partial fulfilment of the requirements for the degree of Bachelor of Science in Fisheries. Prepared at the Department of Aquatic Resources, Ecology, and Management under the supervision of Dr. Ravelina R. Velasco.

## LITERATURE CITED

- Appelbaum, S., Kamler, E. 2000. Survival, growth, metabolism and behaviour of *Clarias gariepinus* (Burchell 1822) early stages under different light conditions. *Aquac. Eng.* 22, 269–287.
- Bagnara, J.T. and M.E. Hadley. 1973. Chromatophores and colour change – The comparative physiology of animal pigmentation. Prentice-Hall, Inc. New Jersey. 202 p.
- Bond, C.E. 1996. *Biology of Fishes*. Saunders College Pub. Philadelphia, USA. p. 750.
- Browman, H.I. and B.M. Marcotte. 1987. Effects of prey color and background color on feeding by Atlantic salmon alevins. *The Progressive Fish-Culturist*, 49(2): 141–143.
- Brummet, R.E. 1995. Environmental regulation of sexual maturation and reproduction in tilapia. *Reviews in Fisheries Science*, 3(3): 231–248.
- Carvalho, T.B., F.Z. Mendonça, R.S. Costa-Ferreira and E. Gonçalves-de-Freitas. 2013. The effect of increased light intensity on the aggressive behavior of the Nile tilapia, *Oreochromis niloticus* (Teleostei: Cichlidae). *Zoologia* 30 (2): 125–129
- Cho, C.Y. and S.J. Kaushik. 1990. Nutritional energetics in fish: energy and protein utilization in Rainbow trout (*Salmo gairdneri*). *World Rev. Nutr. Diet.* 61:132–172.
- Downing, G. and M.K. Litvak 1999. The effect of photoperiod, tank colour and light intensity on growth of larval haddock. *Aquaculture International*, 7: 369–382.
- Duston, J., and R.L. Saunders. 1990. The entrainment role of photoperiod on hypoosmoregulatory and growth-related aspects of smolting in Atlantic salmon (*Salmo salar*). *Canadian Journal of Zoology*, 68(4): 707–715.
- Eames, S.C., L.H. Philipson, V.E. Prince and M.D. Kinkel. 2010. Blood Sugar Measurement in Zebrafish Reveals Dynamics of Glucose Homeostasis. *Zebrafish* 7(2): 205–213.
- Eslamloo, K., S.R. Akhavan, A. Eslamifarz and M.A. Henry. 2013. Effects of background colour on growth performance, skin pigmentation, physiological condition and innate immune responses of Goldfish, *Carassius auratus*. *Aquaculture Research*, 1–14.

- Elnwishy, N., D. Sabri and F. Nwonwu. 2012. The effect of difference in environmental colors on Nile tilapia (*Oreochromis niloticus*) production efficiency. *International Journal of Agriculture & Biology*, 14:516–520.
- Elsbaay, A.M. 2013. Effects of photoperiod and different artificial light colors on Nile tilapia growth rate. *IOSR Journal of Agriculture and Veterinary Science*, 3(3): 5-12.
- Fernandez, P.J. and J.T. Bagnara. 1991. Effect of background color and low temperature on skin color and circulating alpha-MSH in two species of leopard frog. *Gen. Comp. Endocrinol.* 83, 132–141.
- Green, J.A., B.I. Baker and H. Kawauchi. 1991. The effects of rearing rainbow trout on black or white backgrounds on their secretion of melanin-concentrating hormone and their sensitivity to stress. *J. Endocr.*, 128: 267–274.
- Groneveld, D., P.H.M. Balm and S.E. Wendelaar Bonga. 1995. Biphasic effect of MCH on  $\alpha$ -MSH release from the tilapia (*Oreochromis mossambicus*) pituitary. *Peptides*, 16(5): 945-949.
- Gupta, M.V. and B.O. Acosta. 2004. A review of global tilapia farming practices. Retrieved from [www.library.enarca.org](http://www.library.enarca.org) on May 19, 2016.
- Han, D., S.H. Xie, W. Lei, X. Zhu and Y. Yang. 2005. Effect of light intensity on growth, survival and skin of juvenile Chinese longsnout catfish (*Leiocassis longirostris* Günther). *Aquaculture*, 248: 299-306.
- Henne, J.P. and W.O. Watanabe. 2003. Effects of light intensity and salinity on growth, survival, and whole-body osmolality of larval southern flounder (*Paralichthys lethostigma*). *J. World Aquac. Soc.*, 34(4): 450–465.
- Imanpoor, M.R. and M. Abdollahi. 2011. Effects of tank color on growth, stress response and skin color of juvenile Caspian kutum (*Rutilus frisii* Kutum). *Global Veterinaria*, 6(2): 118-125.
- Jentoft, S., S. Øxnevad, A.H. Aastveit and Ø. Andersen. 2006. Effects of tank wall color and up-welling water flow on growth and survival of Eurasian perch larvae (*Perca fluviatilis*). *Journal of the World Aquaculture Society*, 37: 313-317.
- Karakatsouli, N., S.E. Papoutsoglou and G. Manolezios. 2007. Combined effects of rearing density and tank colour on the growth and welfare of juvenile white sea bream *Diplodus sargus* L. in a recirculating water system. *Aquaculture Research*, 38, 1152-1160.

- Lamers, A.E., P.H.M. Balm, H.E.M.G.Haenen, B.G.Jenks and S.E.Wendelaar Bonga. 1991. Regulation of differential release of  $\alpha$ -melanocyte-stimulating hormone forms from the pituitary of a teleost fish, *Oreochromis mossambicus*. J. Endocrinol.129: 179-187.
- Liao I.C. and S.L. Chang, 1983.Studies on the feasibility of red tilapia culture in saline water.Pages 524 - 533 in L. Fishelson and Z. Yaron, compilers.Proceedings of the International Symposium on Tilapia in Aquaculture, Nazareth, Israel, 8 - 13.
- Liao, I.C. and T.P. Chen.1983. Status and prospects of tilapia culture in Taiwan in L. Fishelson and Z. Yaron, compilers. Proceedings of the International Symposium on Tilapia in Aquaculture, Pages 588 – 598 as cited by Watanabe, W.O., D.H. Ernst, B.L. Olla and R.I. Wicklund. 1989. Aquaculture of red Tilapia (*Oreochromis* sp.) in marine environments state of the art. Advances in tropical Aquaculture. Tahiti, pp 487-498.
- Liu, I. 2012. Why Sleeping in Darkness is Essential to Your Health. Retrieved from <http://thenakedlabel.com/blog> on July 19, 2017.
- Llamas, A. H. and V.M.G. Muñoz. 1996. Growth and survival of the Catarina scallop *Argopecten circularis* (Sowerby) to stocking density and length of culture period. Aquaculture research 27: 711 – 719.
- Lovshin, L.L. Undated.Criteria for Selecting Nile Tilapia and Red Tilapia for Culture. Department of Fisheries and Allied Aquacultures. Auburn University, Alabama 36849 USA. 13p.
- Lucas, A. (ed). 1996. Bioenergetics of Aquatic Animals.Taylor& Francis, Ltd. London.67 p.
- Lugert, V., G. Thaller, J. Tetens, C. Schulz and J. Krieter. 2014. A review on fish growth calculation: multiple functions in fish production and their specific application. Reviews in Aquaculture 6, 1–13.
- Mazeaud, M. M., F. Mazeaud and E. M. Donaldson. 1977. Primary and Secondary Effects of Stress in Fish: Some New Data with a General Review. Transactions of the American Fisheries Society. Vol. 106, No.3 201:212.
- Mesa, M.G. and C.B. Schreck. 1989. Electrofishing Mark–Recapture and Depletion Methodologies Evoke Behavioral and Physiological Changes in Cutthroat Trout, Transactions of the American Fisheries Society, 118:6, 644-658
- Ostrowski, A.C. 1989. Effect of rearing tank background color on early survival of dolphin larvae.The Progressive Fish-Cutturist, 51(3):161-163.

- Papoutsoglou, S.E., G. Mylonakis, H. Miliou, N.P. Karakatsouli and S. Chadio. 2000. Effects of background color on growth performances and physiological responses of scaled carp (*Cyprinus carpio* L.) reared in a closed circulated system. *Aquacultural Engineering* 22 309–318.
- Papoutsoglou, S.E., N. Karakatsouli and G. Chiras. 2005. Dietary l-tryptophan and tank colour effects on growth performance of rainbow trout (*Oncorhynchus mykiss*) juveniles reared in a recirculating water system. *Aquacultural Engineering* 32 277–284.
- Patriche, T. 2009. The importance of glucose determination in the blood of the Cyprinids. Morphological Science Dept., Faculty of Medicine and Pharmacy, “Dunarea de Jos” University Galati, Romania. *Lucrări științifice Zootehnie și Biotehnologii*, 42 (2): 102-106.
- Pearson, O. 2017. Is Glucose Stored in the Human Body? Retrieved from <http://www.livestrong.com/article/461651> on September 5, 2017.
- Philippart, J-CL. and J-CL. Ruwet. 1982. Ecology and distribution of tilapias. p.15 – 59. *In: R.S.V. Pullin and R.H. Lowe-McConnell (editors). The Biology and Culture of Tilapias. ICLARM Conference Proceedings 7. International Center for Living Aquatic Resources Management. Manila, Philippines. 432 p.*
- Polakof, S., S. Panserat, J.L. Soengas and T.W. Moon. 2012. Glucose metabolism in fish: a review. *J. Comp. Physiol. B*, 182: 1015-1045.
- Popma, T. and M. Masser. 1999. Tilapia life history and biology. Southern Regional Aquaculture Center. SRAC Publication No. 283. Stoneville, Mississippi. 4p.
- Pottinger, T. G. and Carrick, T.R. 1999. A comparison of plasma glucose and plasma cortisol as selection markers for high and low stress-responsiveness in female rainbow trout. *Aquaculture*, 175 (3-4). 351-363.
- Rakocy, J.E. 1989. Tank culture of tilapia. SRAC Publication No. 282 Aquaculture Centre Southern Region. Texas, pp 4.
- Reid, S.G., N.J. Bernier and S.F. Perry. 1998. The adrenergic stress response in fish: control of catecholamine storage and release. *Comparative Biochemistry and Physiology Part C*, 120: 1-27.
- Rotllant, J., L. Tor, D. Montero, M. Pavlidis, M. Martinez, S.E. Wendelaar Bonga and P.H.M. Balm. 2003. Background colour influence on the stress response in cultured red porgy *Pagrus pagrus*. *Aquaculture*, 223: 129–139.
- Rotllant J., P.H.M. Balm, J.P. Sanchez, S.E. Wendelaar- Bonga Tort L. 2001. Pituitary and interrenal function in gilthead sea bream (*Sparus aurata* L., Teleostei) after

- handling and confinement stress. *General and Comparative Endocrinology* 121, 333–342.
- Rueda, P., Schrama, J.W., Verreth, J.A.J. 2004. Behavioural responses under different feeding methods and light regimes of the African catfish (*Clarias gariepinus*) juveniles. *Aquaculture* 231, 347–359.
- Schreck, C.B., B.L. Olla and M.W. Davis. 1997. Behavioural responses to stress. p. 145-170. *In: Iwama, G.K., Pickering, A.D., Sumpter, J.P., Schreck, C.B. (Eds.). Fish Stress and Health in Aquaculture. Cambridge University Press. Cambridge. 507 p.*
- Tamazouzt, L., B. Chatain and P. Fontaine. 2000. Tank wall colour and light level affect growth and survival of Eurasian perch larvae (*Perca fluviatilis* L.). *Aquaculture*, 182: 85-90.
- van der Salm, A.L., F.A.T. Spanings, R. Gresnigt, S.E. Wendelaar Bonga and G. Flik. 2005). Background adaptation and water acidification affect pigmentation and stress physiology of tilapia, *Oreochromis mossambicus*. *General and Comparative Endocrinology* 144 51–59.
- Velasco, R.R. 2011. Biological stress markers associated with growth of Nile tilapia (*Oreochromis niloticus*) reared at different stocking density. Ph.D. Thesis. Institute of Graduate Studies, Central Luzon State University, Science City of Muñoz, Nueva Ecija, Philippines. 115p. (unpublished)
- Volpato, G.L., C.R.A. Duarte and A.C. Luchiari. 2004. Environmental color affects Nile tilapia reproduction. *Brazilian Journal of Medical and Biological Research*, 37: 479-483.
- Volpato, G.L. and R.E. Barreto. 2001. Environmental blue light prevents stress in the fish Nile tilapia. *Brazilian Journal of Medical and Biological Research*, 34: 1041-1045.
- Watanabe, W.O., D.H. Ernst, B.L. Olla and R.I. Wicklund. 1989. Aquaculture of red Tilapia (*Oreochromis* sp.) in marine environments state of the art. *Advances in Tropical Aquaculture, Aquacop Ifremer Actes de Colloque* 9 pp 487-498.
- Wendelaar Bonga, S.E. 1997. The stress response in fish. *Physiol. Rev.*, 77(3):591-625.
- Zang, L., Y. Shimada, Y. Nishimura, T. Tanaka and N. Nishimura. 2013. A Novel, Reliable Method for Repeated Blood Collection from Aquarium Fish. *The fish haus*. Volume 10: 425-431.

<http://www.nyu.edu/about/news-publications/news/2012/january/advantages-of-the-dark-the-multiple-evolution-events-of-blind-cavefish.html>